

Quantification of rickettsial load during the acute phase of experimental bovine anaplasmosis

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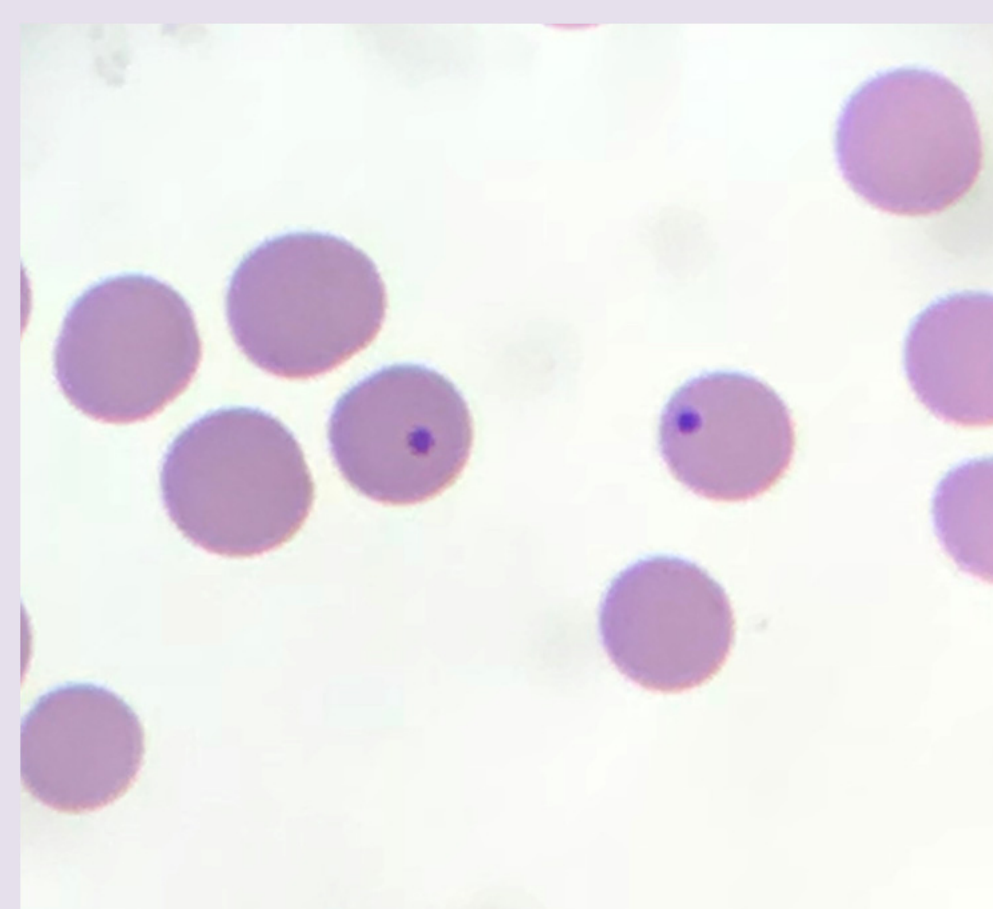
Abstract

Anaplasma marginale is a prototypical tick-borne pathogen, which is the primary etiologic agent of bovine anaplasmosis. The disease has been characterized in depth, with well-established acute and subsequent subclinical phases. Although ascending levels of parasitemia (percent infected erythrocytes) are well established during the incubation through acute phases of bovine anaplasmosis, there are data indicating the highest rickettsial load in peripheral blood of a related pathogen. *Ehrlichia canis*, is not always correlated with the acute phase of canine monocytic ehrlichiosis. This discrepancy could be due to differences in pathogen numbers within infected host cells. Thus, **the central hypothesis of this study is that total rickettsial load in the peripheral blood is directly correlated with parasitemia during the onset of acute bovine anaplasmosis.** Six calves were experimentally infected with *A. marginale*, and peripheral blood was monitored throughout transitions from incubation, acute and subclinical phases of experimental anaplasmosis *marginale*. A related experiment will test the working hypothesis that different anticoagulants (*i.e.*, EDTA, heparin and sodium-citrate) will not affect qPCR after silica-based template isolation. This study is expected to demonstrate that parasitemia is a direct reflection of rickettsial load. Moreover, these results are expected to determine the utility of these methods for *A. marginale* transmission studies.

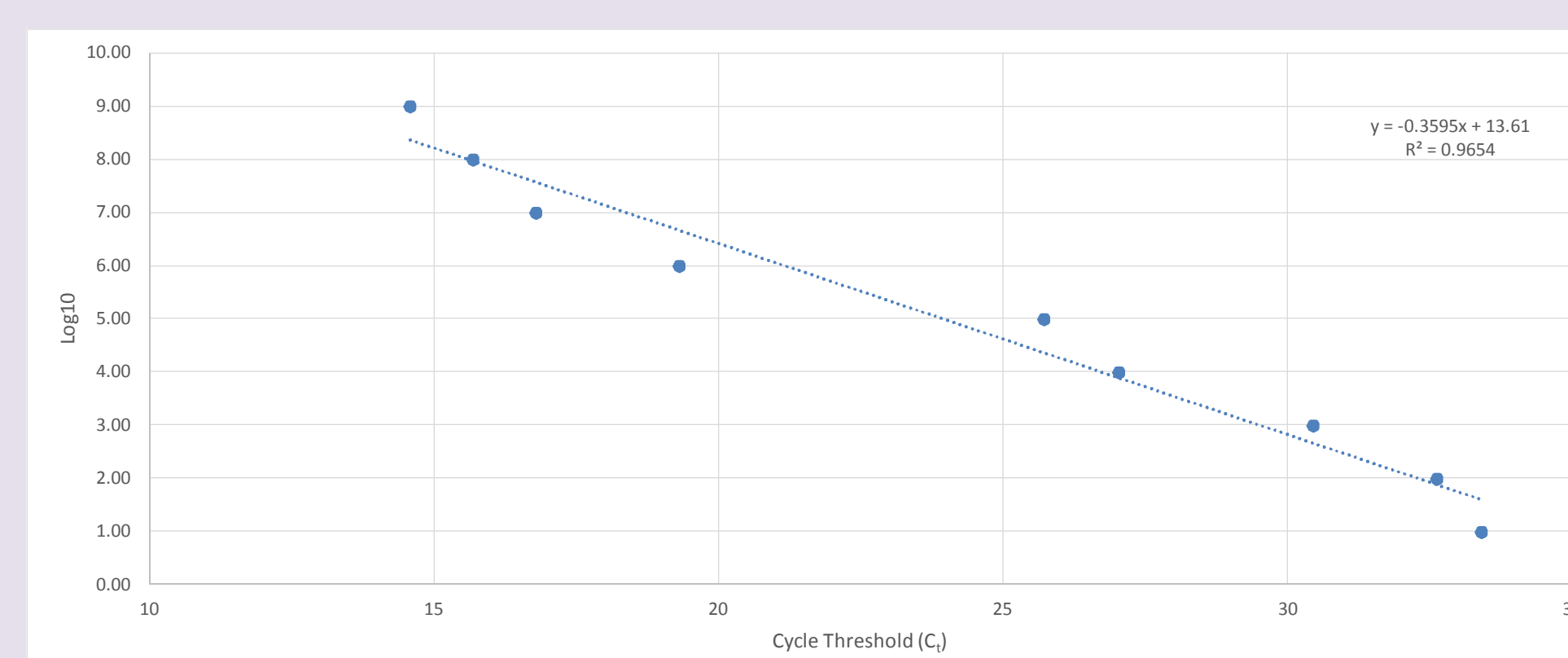
Background

The reason for this experiment was to see how much of a bacterial load was in a cows during the time of infection. A standard curve was produced from a dilution series in order for us to compare the copy numbers we read from the qPCR. As we understood how much bacterial load we had in a cow we then compared it to parasitemia which is shown in **Figure 3.**

Methods

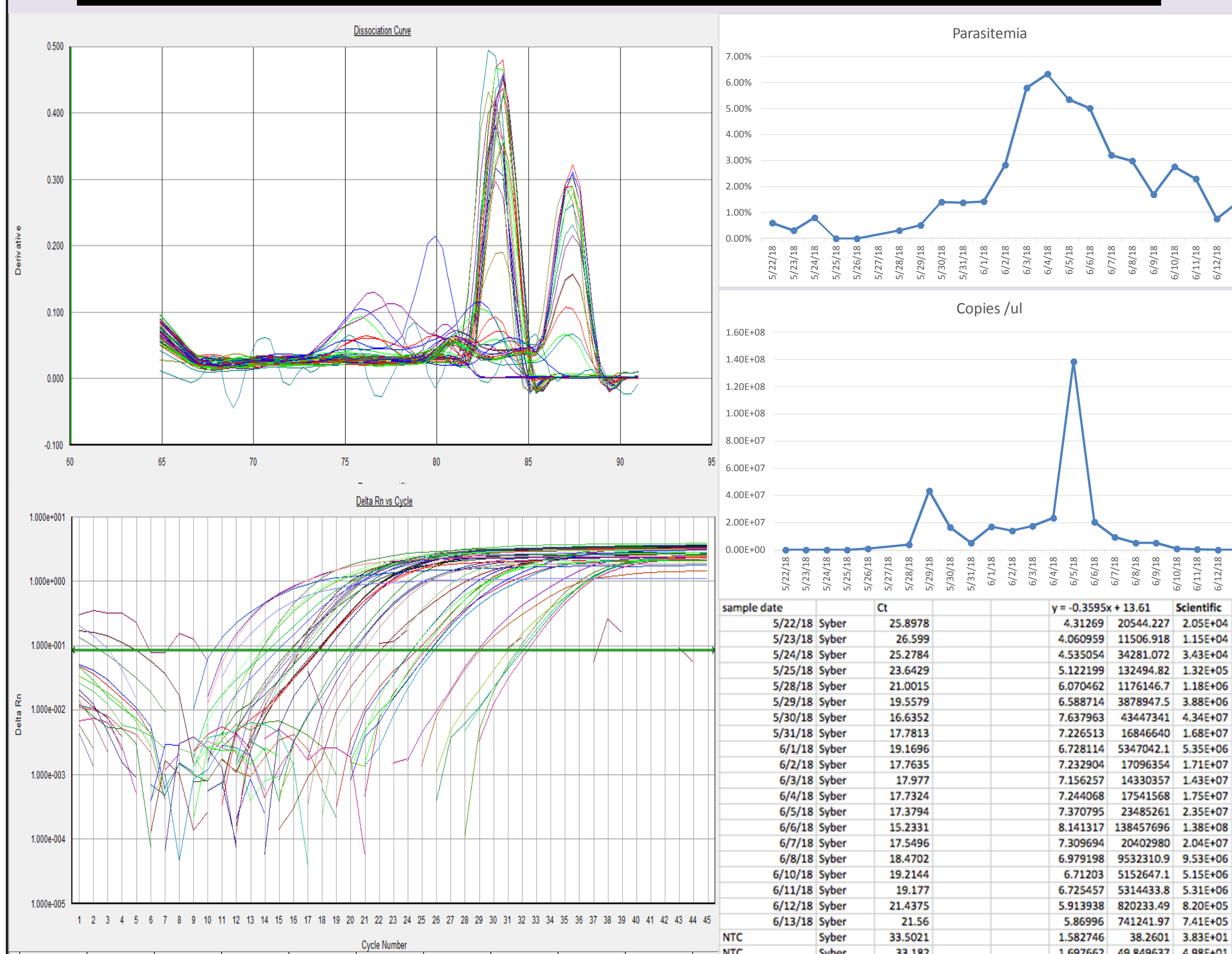


Parasitemias and packed cell volumes were measured with standard methods. Template was isolated with a silica-based spin-column kit (Roche, Indianapolis)(**Figure 1**).



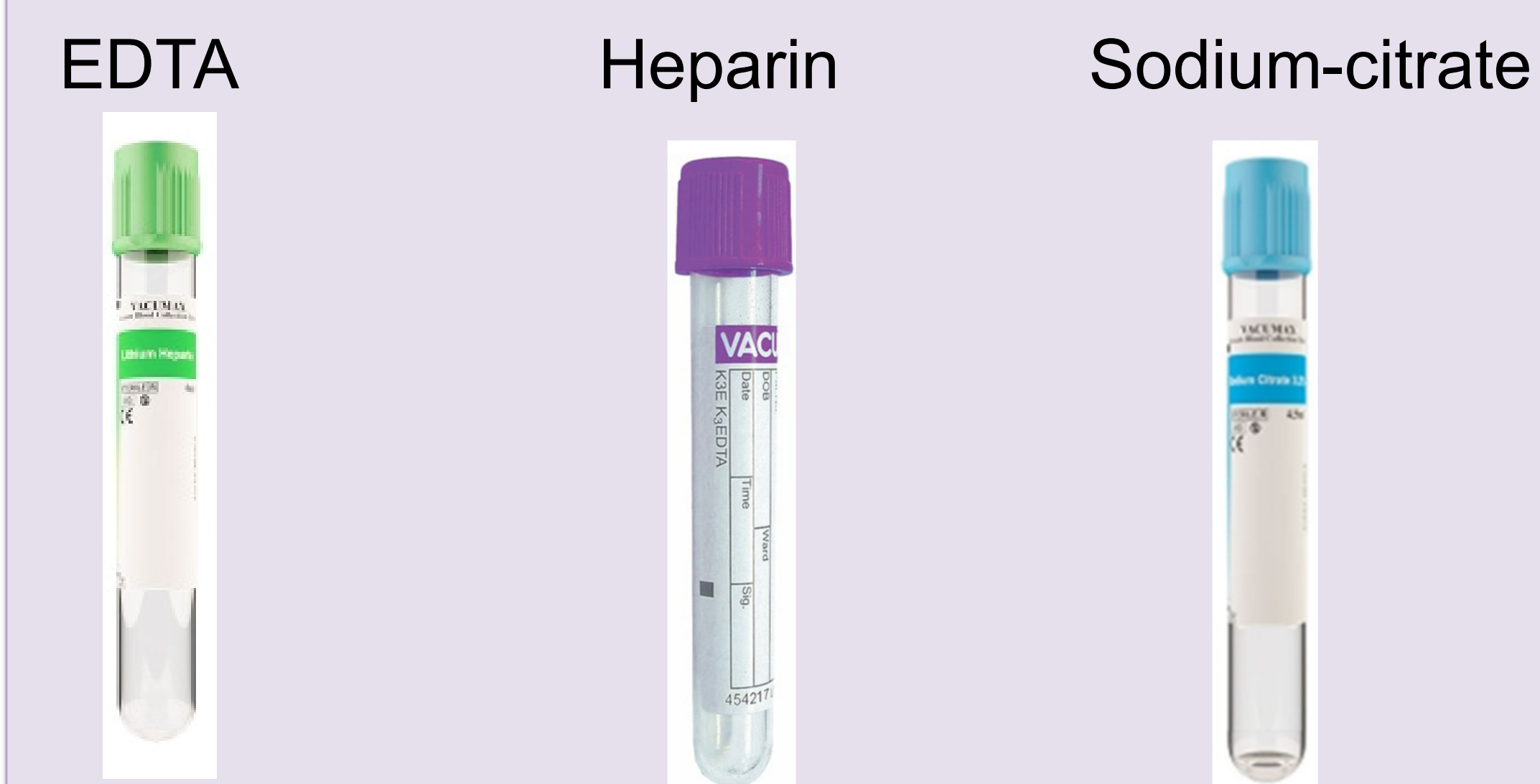
Quantitative PCR(qPCR) will be used to measure rickettsial loads (**Figure 2**).

Quantitative PCR of rickettsial load compared to parasitemia



Cow 2897 was first used in the qPCR experiment. The graph shows the comparison between bacterial load and parasitemia (**Figure 3**)

Different anticoagulants (*i.e.*, EDTA, heparin and sodium-citrate) will not affect qPCR after silica-based template isolation



This experiment will be done at a later date, but these are the tubes we used to extract blood from each cow (**Figure 4**)

Conclusions

In conclusion, cow 2897 was successfully tested with qPCR then compared to its parasitemia and the outcome was that they both correlate to each other.

What's next?

We are working on finding out the hypothesis of different anticoagulants and how they will not affect qPCR after silica-based template isolation

Acknowledgements

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